

MOUNT-QUICK MOUNTING MEDIA PRODUCT #19478

Tissue Transfer Technique:

This technique allows the transfer of tissue or cytology samples from standard slides to positively charged slides. The tissue may be stained or unstained.

Supplies:

- Mount-Quick
- Diamond Marking Pen
- Scalpel blade
- Charged slides
- Graded alcohols
- Xylene

- 1) With the diamond pen, mark the area of interest on the underside of the slide
- 2) A. For stained sections, remove the coverslip by soaking in xylene
B. For unstained sections or smears, dehydrate to xylene in preparation for mounting media.
- 3) Spread the Mount-Quick over the entire area, making sure the slide is coated in xylene. Be sure to form a meniscus over the tissue.
- 4) Place the slide in a 60°C oven for at least 1.5 to 2 hours or a 37°C oven overnight until the mounting media hardens. To continue—the media **MUST** be hardened.
- 5) With the diamond pen, mark the area on the mounting media surface to the corresponding area on the underside of the slide.
- 6) Soak the slide in warm water for at least an hour or more.
- 7) Test the release of the media by trying to pry the edges up with a scalpel blade. If the mounting media does not peel off easily, continue soaking.
- 8) Once the section is removed, place it on a charged or adhesive coated slide. Make certain the section is placed with the same side down as on the original slide.
- 9) Wet the slide and the media so they will adhere.
- 10) With gloves on, take a gauze soaked in warm water and place it on top of the section. With your thumb, press the gauze flat and express out all the water. This action will assist in keeping the edges of the media down.
- 11) Place slide in an oven (37-60°C) in a horizontal position for one hour or longer.
- 12) Once the slide is dry and adhered, begin the process of media removal.
- 13) Place the slide in xylene (4 changes, 3 minutes each) until all Mount-Quick is dissolved. If needed, add 2-4 minutes to each xylene
- 14) Rehydrate through ethanol (2 changes each) 100%, 95% and water.

Notes:

Do not use temperatures higher than 60°

The best adhesion is achieved with positively charged adhesive slides

If a hot plate was used to spread the paraffin section after the original sectioning, removal of the entire section is VERY difficult. Extended soaking of the section does not appear to improve the results.

REFERENCES:

“ The Preparation of Serial Microscopic Sections in Form of Plastic Films.” Demonstration at the Annual meeting of the International Academy of Pathology, Weibel, E., Shenk, R., Morger, R., Toendury, G., Boston, 1959

“ Application of Diatex Compound in Cytology: Use in Preparing Multiple Slides from a Single Routine Smear”, Jimenez, Joseph D., Gangi, MD., Acta Cytol 30:445, 1986.

“ Diagnostic Immunocytochemistry and Electron Microscopy-Preparing Multiple Slides from a Single Smear”, Yazdi, H., Dardick, I., Igaku-Shoin, New York-Tokyo, 1992, pp. 36-39.